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Leucas Aspera to Treat Pan Oral Disorders - An Experimental Study

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Abstract

Objectives: This article aimed at discussing the effectiveness of developing a new antioxidant and anti-inflammatory product from the natural source namely the extract of Leucas aspera plant that can be used to treat various oral mucosal conditions.

Study design: The study was a descriptive experimental study. The dried powder extract of the whole LA plant was subjected to various procedures such as phytochemical analysis, antioxidant assay (DPPH) and assessment of anti-inflammatory effect (Cytokine activity in RAW 264.7 macrophage cells) and the results were tabulated. Significance of anti-inflammatory activity was tested by T-test.

Results: Our analysis has confirmed that $100\mu g$ of plant extract showed high amount of antioxidant activity (78%). The cytokine inhibitory percentage of our plant extract showed consistent increase when tested from 250 ug/ml to 1000u g/ml to be from 24% to 39% thus demonstrating significantly high level of anti-inflammatory potential (P > 0.01)

Conclusion: Our study suggests that LA plant can be safely utilized for testing and developing products based on its extract.



Article History

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Keywords

Antioxidants; Anti-Inflammatory; Efficiency; Leucas.

Introduction

The existence of widespread nutritional deficiencies and tobacco related habits makes occurrence of aphthous ulcers and premalignant lesions such as oral leukoplakia, erythroplakia, lichen planus, actinic cheilitis, submucous fibrosis as well as palatal lesions in reverse smokers are more common among the Indian population. The discovery of medicinal plants and their usage in drug manufacturing has attracted many pharmacological companies owing



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to their potent pharmacological activities, lower toxicities and economic viability.1 The roots and stems of leucas aspera (LA) has demonstrated high antioxidant activity2 in comparison to vitamin E and thus can be included as an active formula in treating such potentially malignant disorders. The anti-inflammatory property of leucas aspera would also be beneficial in treatment of aphthous ulcers and stomatitis. Based on the outcome of phytochemical properties, animal studies³ and culture analysis; plant extract showed high amount of antioxidant activity. Leucas is an ideal extract to treat the most common oral disorders as a whole; based on its antioxidant and anti-inflammatory efficiency. It can be considered as a potential substrate to develop all-purpose medication for oral topical usage.

The presence of highly potent pharmacological activities, lower toxicities and economic viabilities have sorted many pharmacological companies to utilize medicinal plant for preparing drugs. These indigenous medicinal plants have opened a new arena

Table 1: Phytochemical Analysis of herbal extract

| Chemical test - Extract | Presence/ Absence |
|-----------------------------|----------------------|
| Carbohydrate | |
| Fehling's test | ++++ |
| Molisch test | ++++ |
| Alkaloids | |
| Dragendroff's test | ++ |
| Wagner's test | ++ |
| Hagers's test | +++ |
| Mayer's test | ++++ |
| Glycosides | |
| Keller Killani test | ++++ |
| Saponins | |
| Foam test | ++++ |
| Phenolic compounds | |
| and tannin | |
| Ferric chloride test | ++++ |
| Test for Sterols | Negative (-) |
| (++) = Moderate content | |
| (+++) = substantial content | |
| (++++) =high content | |
| (-) = Absent. | |

for drug development. The presence of antioxidant properties in addition to antimicrobial properties has kindled new interests in the development of these plant-based products on premalignant lesions and conditions of the oral cavity.

Various commonly occurring oral lesions do not have a simple topical pharmacological management and require per-oral medications or invasive procedures such as biopsies for their effective treatment. Major lesions of importance are as follows:

Oral Squamous Cell Carcinomas (OSCC) are principally induced by free radicals such as reactive oxygen and nitrogen species (ROS and RNS) through oxidative and nitrative stress.⁴ The free radicals as well as protein damage play key roles in carcinogenesis as they cause DNA base alterations, strand breaks, damaged tumor suppressor genes, and proto-oncogenes expression. RNS occurs in the form of ROS such as superoxide radicals (O₂), hydroxyl radicals (OH), nitrosamines (NO₃ and NO₂); and hydrogen peroxide (H₂O₂).⁴

Recurrent Aphthous Stomatitis (RAS)

Recurrent, multiple, small, round, or ovoid ulcers with circumscribed margins, having yellow or gray floors. This is a relatively more common painful condition among various populations. The ulcers first appear during childhood or adolescence and are marked by erythematous haloes surrounding them.⁵

Oral Submucous Fibrosis (OSMF) is one of oral potentially malignant disorders owing to the habit of chewing areca nut. It is a chronic condition of insidious origin and can affect all parts of oral cavity including pharynx. It can also be associated with vesicle formation that may precede the lesion proper sometimes. Histology the condition can be defined as having a juxta-epithelial inflammatory reaction followed by a fibroelastic change of the lamina propria, with epithelial atrophy leading to stiffness of the oral mucosa. It causes trismus and inability to eat as a clinical manifestation.⁶

Oral Leukoplakia (OL) has been defined by World Health Organization (1978) "as a white patch or plaque which cannot otherwise be characterized clinically or pathologically as any other disease".

The other potentially malignant disorders are palatal lesions in reverse smokers, erythroplakia, actinic cheilitis and lichen planus.⁷ Erythroplakia and leukoplakia are far more common lesions in tobacco users. The malignant transformation potential of leukoplakia is 0.13% to 17.5% and OSMF is 2-8%7,8. The current protocol in the management of potential malignant disorders is habit intervention followed by active treatment in case of erythroplakia as well as leukoplakia. The non-surgical management of leukoplakia includes treating the lesions using carotenoids, vitamins, bleomycin and photodynamic therapy. The more accessible and circumscribed lesions were treated with excisional biopsy.

Materials and Methods Preparation of Plant Extract

The leaves, stem, roots and flowers of the plant were sundried for 1 week and were ground into fine powder using regular food processor. The obtained powder was soaked in 500ml methanol (80%v/v) and kept at room temperature (28-30 degree centigrade) for 4 days. The Whatman's filter paper was used to filter the extract and the obtained fluid was utilized for various analysis.

Phytochemical Analysis

The obtained extract was run through various phytochemical analysis and the results were tabulated. 10mg of plant extract is dissolved in 10ml of methanol(1mg/ml) and 500ul extract was used for each reaction. Tests for carbohydrates, flavonoids, sterols, alkaloids, glycosides, saponin, phenolic compounds and sterols were done.

Antioxidant Assay

The extract was also analyzed for the presence of antioxidant activity which showed significant scavenging action.

Anti-Oxidant Activity

2, 2-diphenyl-1-picrylhydrazyl (DPPH) assay:

The colorimetric DPPH assay by Shimada *et al.*, (1992)⁸ was utilized to determine the radical scavenging activity of the plant extracts.

Principle of the Assay

The assay is based on the hydrogen donating capacity of test samples. This capacity is quantified in terms of their ability to scavenge the relatively stable, organic free radical DPPH and by their consequent reduction. The DPPH solution colored deep violet was measured at 550nm. In the event of subsequent reduction; decolorization to yellow-white occurs. The degree of reduction can be measured stoichiometrically. The radical scavenging activity of sample extract will correspond inversely to DPPH measured earlier at 550nm.8

Spectrophotometric Determination of the Antioxidant Activity of the Plant Extract

The radical scavenging activity of the extract at different concentrations prepared using ethanol was obtained after utilizing a 96-well microtiter plate to generate the quantitative measure. The methanol extract with different concentrations were prepared. DPPH solution was prepared in ethanol. Aliquots of plant extract prepared in ethanol is added to DPPH solution. This plate was shaken

| Table 2: DPPH assay - | Absorbance of | the extreete | at EOEnm |
|-----------------------|---------------|--------------|----------|
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| Volume taken from extract (ul) | Amount of extract- Content(ug) | Absorbance (OD) at 595nm | % scavenging activity |
|--------------------------------|-----------------------------------|-----------------------------|-----------------------|
| Blank(methanol) | | 0.9988 | |
| 0.1ul | 0.1ug | 0.7861 | 21.17 |
| 0.5ul | 0.5ug | 0.4462 | 55.2 |
| 1ul | 1ug | 0.3620 | 63.63 |
| 10ul | 10ug | 0.2944 | 70.40 |
| 30ul | 30ug | 0.2536 | 74.48 |
| 100ul | 100ug | 0.2115 | 78.70 |

mildly for 2 min, after which it was stored in the dark for 30 min. The microplate reader was used to measure the decolorization spectroscopically at 595nm. The negative controls contained plant extract to methanol was added, in the absence of DPPH. Percentage purple decolorization to DPPH yellow was then determined for samples as a measure of the free radical scavenging activity. Lower absorbance of the reaction mixture indicated higher free radical scavenging activity. % DPPH radical-scavenging = (Absorbance of control-Absorbance of test Sample) X100 (Absorbance Of control)⁸

Anti-Inflammatory Properties of LA

In addition to various other functions of IL-1 β ; it also is an inflammatory mediator and is involved in a variety of cellular activities, including cell proliferation, differentiation, and apoptosis. IL-1 β which will be produced by activated macrophages is proteolytically processed to its active form by caspase 1 (CASP1/ICE). The Central Nervous System (CNS) activation through cyclooxygenase-2 (PTGS2/COX2) by this cytokine is found to contribute to inflammatory pain hypersensitivity. The LA extract was tested for the effective inhibition of the IL-1 β obtained from RAW 264.7 macrophages.

Cell Culture

Mouse macrophage cell line RAW 264.7 obtained from National Center for Cell Science (Pune, India) was utilized for the purpose. The RAW 264.7 macrophages were cultured in DMEM medium. Dulbecco's Modified Eagle's Medium (DMEM)) containing 10% fetal bovine serum (FBS) supplemented with antibiotics solution was used

to culture the cells. The cells were grown at 37°C and 5% CO₂ – 95% air under humidified conditions. A 96-well microplate was used for culturing RAW 264.7 macrophages at an initial density of 1 × 104 cells/well for 24 h. The cells were subsequently treated with plant extract at various concentrations (250, 500, or 1000 μg/mL) and LPS (1 μg/mL) for additional 24 hours. LPS was dissolved in DMEM. The cells were stimulated with plant extract and LPS (1 µg/mL) for 24 h to assess the effect of plant extract on the production of IL-1 in the LPS-induced RAW 264.7 macrophages and the cell culture supernatants were collected. An ELISA kit was used to measure the IL-1β content of the supernatants according to the manufacturer's protocol and absorbances (nm) were also recorded using a microplate reader.8

Preparation of Samples for ELISA

The kit used was Human osteocalcin ELISA Kit, Catalogue No.: EH3468 from Fn-Test China;Range: 1.25-80ng/ml, Sensitivity: < 0.75ng/ml. The sample preparation was done by cell lysate. Trypsin was utilized to detach adherent cells. 1)cell suspensions were collected by direct centrifugation. 2) cells were washed thrice in PBS. 3) cells were re-suspended in PBS and subjected to lysis using RIPA buffer and frozen cells at -20°C. 4) cells were thawed. 5) Centrifuged at 1000×g (or 3000 rpm) for 15 minutes at 2-8°C to remove cellular debris. 6) Samples stored at -20°C.8

Cytokine Activity of the LA in RAW 264.7 Macrophage Cells

Enzyme-linked immunosorbant assay (ELISA) technique was carried out using simultaneous profiling of the level of multiple cytokines and/or

| _ | pg/ml (IL-1Beta levels) | | | | |
|-----------------------------|--|--|---|--|--|
| Details | 1 | 2 | 3 | Cytokine levels(pg/ml) | |
| ug/ml 250 500 1000 | 1000 726.8312 706.1918 602.9947 | 1000 651.8812 672.0792 561.5842 | 1000 788.961 707.5758 589.1775 | 1000 717.3733 689.0383 574.7673 | % Inhibition 23.85291 26.86059 38.99012 |
| LPS(1ug/ml) media | 937.2724 109.2675 | 968.3168 122.7723 | 913.4199 92.64069 | 942.0889 108.3764 | |

Table 3: Cytokine inhibitory effect of the LA extract

chemokines. This was carried out in conventional and simple sandwich-based ELISA. The target-specific capture antibodies were used to coat the 96-well ELISA microplate; one in each eight-well strip. Profiling results from up to six samples was obtained. Reagent included the corresponding detection antibodies, Antigen Standards, and a complete set of detection reagents for a colorimetric ELISA.8

Assay Protocol

The capture antibodies were incubated to bind their specific protein of interest. The unbound protein was washed away. The captured analyte in the wells was treated with biotinylated detection antibodies. The wells were washed to remove unbound material. An avidin-horseradish peroxidase conjugate was added. The wells were again washed and a colorimetric substrate solution was added, which produced a blue color in direct proportion to the amount of protein analyte present in the initial sample. The stop solution was added to stop color development. The absorbance at 450 nm was read and compared across samples. All wells of an eight-well strip were coated with the same capture antibody for the same cytokine or chemokine.⁸

The procedure followed include all reagents to be prepared. The samples, positive and negative controls were set up. A 50 µl assay buffer was added into each well of ELISA Array plate. Compound treatment done using 1000ug/ml, 500ug/ml and 250ug/ml of plant extract. The samples and control samples 50 µl each were transferred into the appropriate wells of the ELISA Array plate and incubated for 2 hours. After three washes; 100µl of Detection Antibody solution was added and incubated for 1 hour. After three washes 100µl Avidin-HRP added and incubated for 30 minutes. The 100µl development solution was added after 4 washes and incubated for 15 minutes in the dark.

Finally; 100µl of stop solution was added and read at OD450nm within 30 minutes. A standard curve was plotted relating the intensity of the color (O.D.) to the concentration of standards. The OD concentration in each sample was interpolated from the standard curve obtained.⁸

Results

The plant extract subjected to various procedures such as phytochemical analysis, Antioxidant assay (DPPH) and Anti-Inflammatory properties assay (Cytokine activity in RAW 264.7 macrophage cells) have been reported as follows:

Phytochemical Analysis

The 500ul plant extract was subjected to tests for carbohydrates, flavonoids, sterols, alkaloids, glycosides, saponin, phenolic compounds and sterols. The results are tabulated in Table 1. The given plant extract from LA showed high amounts of carbohydrates, glycosides and saponins. Significant amounts of alkaloids and Phenolic compounds. There were no sterols isolated from the extract.

Antioxidant Capacity using DPPH Assay

The free radical scavenging activity of LA extract revealed the concentration with maximum antioxidant capacity as tabulated in Table 2. The percentage of scavenging capacity was obtained through the following formula and the reading was done at OD 595nm

% DPPH radical-scavenging = (Absorbance of control -Absorbance of test Sample/ (Absorbance Of control)) X100

The percentage of scavenging activity crossed 50 when the concentration of 0.5ml of extract was added and continued to increase to 63.63, 70.40, 74.48 percent at 1ul, 10ul, 30ul and reached 78% at 100ml

Table 4: Anti-inflammatory activity of LA extract at various concentrations and its significant cytokine inhibitory effect

| Plant extract concentration | Absorbance Mean | Mean Cytokine Level | SD | t value | P value |
|-----------------------------|--------------------|------------------------|--------|---------|---------|
| ug_250 | 0.333 | 717.37 | 317.22 | 5.254 | .003 |
| ug_500 | 0.347 | 689.04 | 319.10 | 5.088 | .004 |
| ug_1000 | 0.484 | 574.77 | 323.34 | 5.166 | .004 |

concentration. The combined extract of the LA plant thus has highly significant antioxidant capacity as indicated in Figure 1.

Cytokine activity in RAW 264.7 macrophage cells: Compound treatment done using 1000ug/ml, 500ug/ml and 250ug/ml of plant extract on Multi-Analyte ELISA Array Kit for Cytokine activity of the LA in RAW 264.7 macrophage cells on 96 well ELISA microplate showed the results as per Table 3. The interleukin (IL-1 β) level and its inhibition was assessed as an indicator of anti-inflammatory activity of the LA extract (Figure 2). The results were analyzed using t test for statistical significance.

Discussion

The phytochemical analysis of LA reveals that its high content of carbohydrates, alkaloids, saponins, Phenols and tannins indicating high potential form medicinal value.

The antioxidant and scavenging ability of phenolic compounds against free radicals is a well-established factor. 10,11 A study proves that phenolics expressed major antioxidant activity of the selected edible flowers. 10 Do *et al.*, revealed that naturally occurring polyphenolics showed the highest antioxidant capacity when compared to synthetic alternatives. 12 The high content of Phenols and Tannins in the LA plant extract (Table 1) reveals that it has sufficient amounts of antioxidant activity being a natural source.

DPPH assay can be used to measure the free radical scavenging action of a compound.¹³ DPPH is a stable free radical that can accept an electron or hydrogen radical to become a stable diamagnetic molecule. DPPH is of deep purple colour with absorption maximum at 517 nm.¹⁴ Antioxidants present in an extract changes DPPH to a colourless compound (2,2-dipenyl-1-picryl hydrazine) due to quenching of

DPPH Assay using LA extract 100 scavenging activity of LA extract 90 80 63,63 70 55.2 50 40 30 21-17 20 10 0 0.1ug 0.5ug 1ug 10ug 30ug 100ug 0.1ul 0.5ul 100ul 1ul 10ul 30ul LA extract concentrations

Fig. 1: DPPH assay measuring the antioxidant capacity of the LA extract

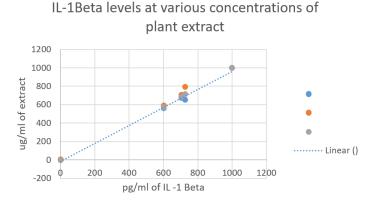


Fig. 2: IL-1 Beta levels plotted against of various concentrations of LA extract

DPPH free radicals and hence used as a substance to evaluate the antioxidant activity. 15,16 In our study, the plant extracts had significant scavenging effects on the DPPH radical which increased with the increase in the concentration of the sample. The 100ug extract demonstrated 78% antioxidant activity when 0.5ug itself demonstrated 55% antioxidant activity proving the extract will be a significantly powerful anti-oxidant (Table 2, Figure 1). Further, the antioxidant activity increased consistently on increasing the concentration of plant extract from 1ul, 10ul, 30ul and 100ul to show 63.63, 70.40, 74.48 and 78.70% scavenging ability thus proving the effectiveness of the plant extract (Figure 1). While the synthetic analogue would cost more; the relatively inexpensive LA plant extract will be able to provide a readily available free radicle scavenge without any adverse effects. High content of tannins also provide credence to local usage of such compound in oxidative stress induced ailments.11

Anti-inflammatory effects in LPS stimulated RAW264.7 cells were studied using ELISA. Any Infection or irritation results in inflammation as the first response. Inflammation is the result of cytokine activity such as TNF-α, IL-1, IL-6, and by eicosanoid such as PGE2.17 LPS is well known to activate two following downstream pathways: the MyD88 and TRIF-dependent pathways.18 The activation of caspase-1 enabled by LPS and ATP stimulation induced robust release of IL-1\beta. IL-1\beta is an important pro-inflammatory cytokine in cancer formation. In addition to many immune reactions; mature IL-1 β is involved in the recruitment of inflammatory cells to the site of infection 19. Monocytes, macrophages and accessory cells secrete IL-1\beta as a soluble protein. These are involved in the activation of T-lymphocytes and B-lymphocytes. Lymphocyte response to antigens or mitogens is mediated by IL-1 β and is released early in an immune response20. Interleukin-1 beta can be activated by a second signaling mechanism involving the inflammasome and caspase 1. The rate-limiting step in the release of IL-1 β is the translation of mRNA into the IL-1β precursor.¹⁰ Active form of monocytes and caspase-1 are already present in circulation. When the monocyte is stimulated to synthesize the IL-1β precursor; cleavage of the precursor takes place and mature IL-1 β is secreted over several hours21. IL-1β, also called endogenous pyrogen, has ability to high level regulation both transcriptionally and post-transcriptionally. It can be induced transcriptionally through number of pathways including through Toll-like receptors (TLRs), which may be activated by microbial pathogens, or by cytokines including TNF- α (tumor necrosis factor- α) and as well as IL-1ß itself. These pathways lead to activation of transcription factors, particularly NF-κB.21

The Cytokine inhibitory percentage of our plant extract showed consistent increase when tested from 250 ug/ml to 1000ug/ml to be from 24% to 39% (Table 3,4, Figure 2) thus demonstrating significantly high level of anti-inflammatory potential (P > 0.01).

Conclusion

Based on these findings there is concrete evidence that Leucas aspera might be considered as a wonder drug to treat many stress induced oral conditions which are on the rise in the modern world. This requires further investigative studies before being able to be clinically used.

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Conflict of Interest

Nil

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